

A Novel Osmotic Pressure Sensor for Interstitial Glucose Assessment: Results from a Laboratory Interference Feasibility Test

Andreas Pfütznner^{1,2}, Rita Smajda³, Guy Voirin³, Gaëlle Andreatta³, Sanja Ramljak², Rune Frisvold⁴, Martha Liley³

¹Sciema – Science & Marketing, Mainz, Germany, ² Pfütznner Science & Health Institute, Mainz, Germany, ³CSEM, Neufchatel, Switzerland, ⁴Lifecare, Bergen, Norway

Introduction

It has been shown that an osmotic pressure sensor based on a lectin/Concanvalin A binding reaction is capable to track glucose changes in an outside environment. An implantable interstitial fluid sensor is currently under development by Lifecare, Bergen, Norway (Sencell). A working laboratory sensor prototype model was used for chemistry optimization experiments and further developmental tasks. One goal was to evaluate the stability of the glucose sensor signal against other interfering substances.

Methods

A working laboratory model of the core sensor technology was developed to perform the interference experiments. For each test, the sensor was equilibrated with 2 mmol/l glucose prior to addition of the potentially interfering substances in high physiological concentrations. The following carbohydrates and concentrations were tested: α -D-Mannose (100 μ mol/l), xylose (1.5 mmol/l), maltose (10.5 mmol/l), fructose (120 μ mol/l), transferrin (5g/L). In case of a measurable impact of the carbohydrate on the sensor signal, at least one more carbohydrate concentration below the initial concentration was to be tested.

Results

No impact on the sensor signal was seen by mannose, xylose, fructose, and transferrin in the tested concentration. A detectable interference was observed with maltose at concentrations of 10.5 mmol/l and 5 mmol/l. With 5 mmol/l maltose, the observed increase in the osmotic pressure signal amounted to ~9 % (10.5 mmol/l: ~17 %). However, such maltose concentrations are unlikely to be seen in the interstitial fluid under physiological conditions in human subjects.

Fig.1: Pictures of the laboratory working pressure cell chamber



Fig.2: Addition of mannose in a concentration of 100 μ M did not influence the sensor signal

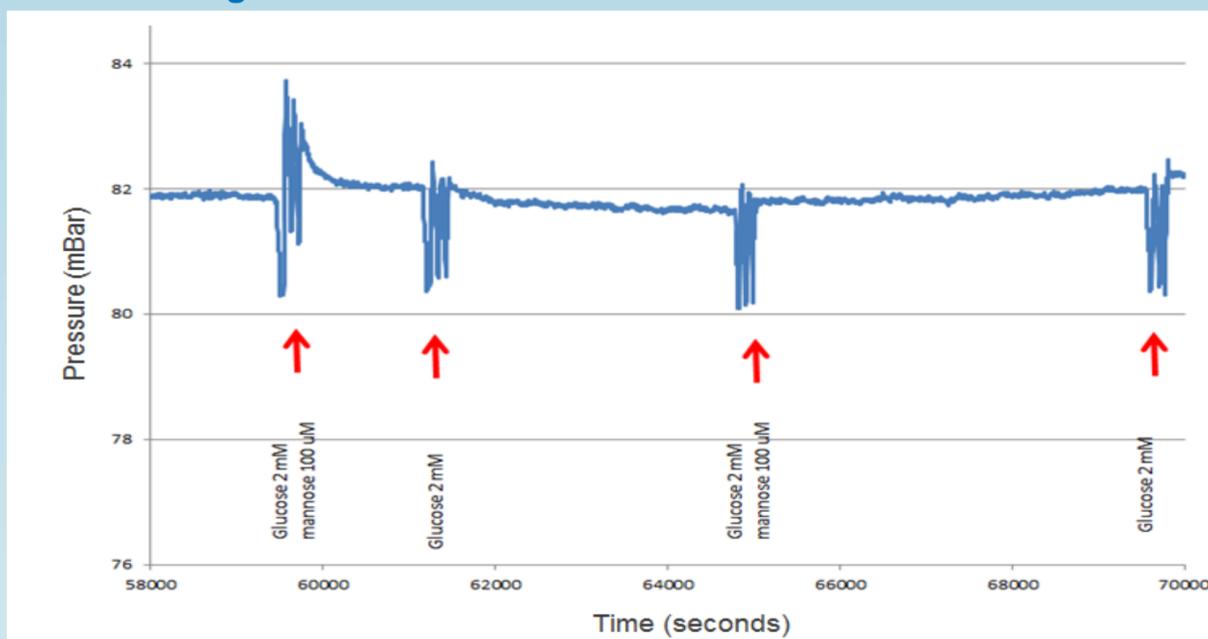
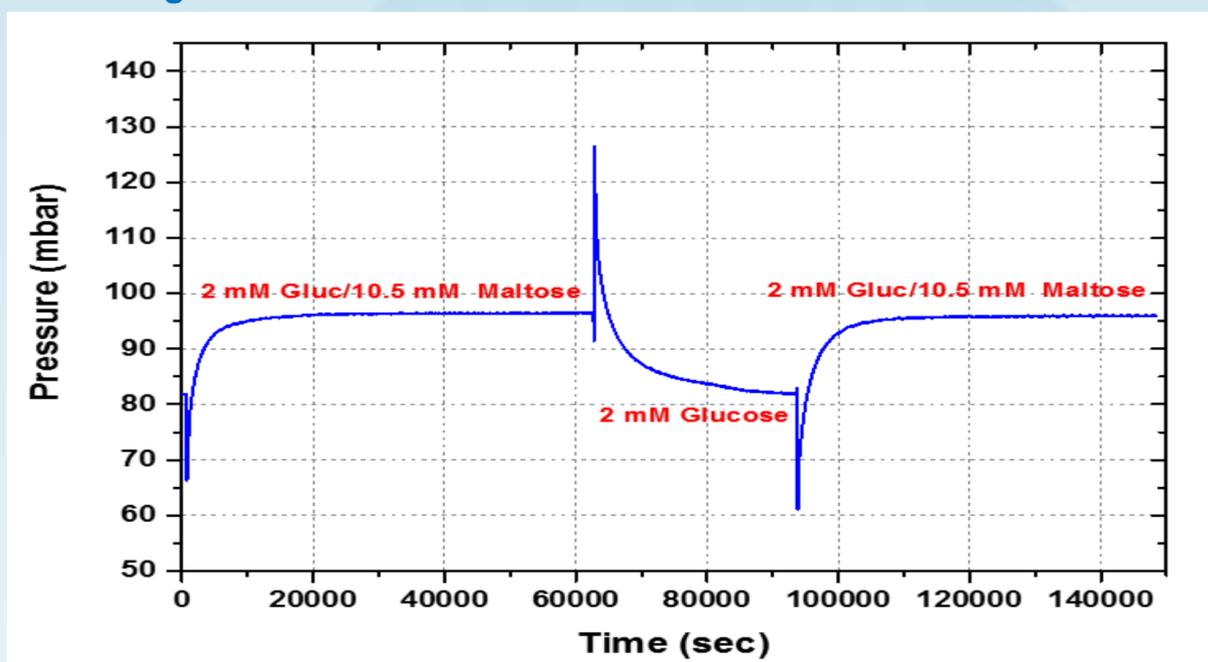


Fig.3: Addition of 10.5 mM maltose had an additive effect to glucose on the sensor signal



Conclusions

The results of our pilot experiments show that the osmotic sensor technology employing the competitive binding of glucose to Concanvalin A vs. dextrane in a core sensor chamber is not influenced by the majority of the tested potentially interfering carbohydrate molecules. The only interfering carbohydrate was maltose in high supra-physiological concentrations. Further tests with the final sensor product will have to confirm these results in a clinical or laboratory setting.